

RESEARCH ARTICLE

# Effects of ocean acidification on primary production in a coastal North Sea phytoplankton community

Tim Eberlein<sup>1\*</sup>, Sylke Wohlrab<sup>1</sup>, Björn Rost<sup>1</sup>, Uwe John<sup>1,2</sup>, Lennart T. Bach<sup>3</sup>, Ulf Riebesell<sup>3</sup>, Dedmer B. Van de Waal<sup>1,4</sup>

**1** Alfred Wegener Institute, Helmholtz Centre for Polar and Marine Research, Am Handelshafen 12, Bremerhaven, Germany, **2** Helmholtz Institute for Functional Marine Biodiversity Oldenburg (HIFMB), Carl von Ossietzky Straße, Oldenburg Germany, **3** GEOMAR Helmholtz Centre for Ocean Research Kiel, Düsternbrooker Weg 20, Kiel, Germany, **4** Netherlands Institute of Ecology (NIOO-KNAW), Wageningen, The Netherlands

\* [tim.eberlein@rub.de](mailto:tim.eberlein@rub.de)



## OPEN ACCESS

**Citation:** Eberlein T, Wohlrab S, Rost B, John U, Bach LT, Riebesell U, et al. (2017) Effects of ocean acidification on primary production in a coastal North Sea phytoplankton community. PLoS ONE 12(3): e0172594. doi:10.1371/journal.pone.0172594

**Editor:** Kay C. Vopel, Auckland University of Technology, NEW ZEALAND

**Received:** September 29, 2016

**Accepted:** February 7, 2017

**Published:** March 8, 2017

**Copyright:** © 2017 Eberlein et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** All relevant data are within the paper and its Supporting Information files.

**Funding:** This project was funded by the German Federal Ministry of Science and Education (BMBF) in the framework of the BIOACID II project (grant no. FKZ 03F0655A). SW, TE and DBvW received support from the EU FP7 research infrastructure initiative 'Association of European Marine Biological Laboratories' (ASSEMBLE (grant no. 227799), and SW from the Royal Swedish Academy of Sciences.

## Abstract

We studied the effect of ocean acidification (OA) on a coastal North Sea plankton community in a long-term mesocosm CO<sub>2</sub>-enrichment experiment (BIOACID II long-term mesocosm study). From March to July 2013, 10 mesocosms of 19 m length with a volume of 47.5 to 55.9 m<sup>3</sup> were deployed in the Gullmar Fjord, Sweden. CO<sub>2</sub> concentrations were enriched in five mesocosms to reach average CO<sub>2</sub> partial pressures (*p*CO<sub>2</sub>) of 760 μatm. The remaining five mesocosms were used as control at ambient *p*CO<sub>2</sub> of 380 μatm. Our paper is part of a PLOS collection on this long-term mesocosm experiment. Here, we here tested the effect of OA on total primary production (PP<sub>T</sub>) by performing <sup>14</sup>C-based bottle incubations for 24 h. Furthermore, photoacclimation was assessed by conducting <sup>14</sup>C-based photosynthesis-irradiance response (P/I) curves. Changes in chlorophyll *a* concentrations over time were reflected in the development of PP<sub>T</sub>, and showed higher phytoplankton biomass build-up under OA. We observed two subsequent phytoplankton blooms in all mesocosms, with peaks in PP<sub>T</sub> around day 33 and day 56. OA had no significant effect on PP<sub>T</sub>, except for a marginal increase during the second phytoplankton bloom when inorganic nutrients were already depleted. Maximum light use efficiencies and light saturation indices calculated from the P/I curves changed simultaneously in all mesocosms, and suggest that OA did not alter phytoplankton photoacclimation. Despite large variability in time-integrated productivity estimates among replicates, our overall results indicate that coastal phytoplankton communities can be affected by OA at certain times of the seasonal succession with potential consequences for ecosystem functioning.

## Introduction

Atmospheric CO<sub>2</sub> partial pressure (*p*CO<sub>2</sub>) is currently rising at an unprecedented rate due to anthropogenic activities. This leads to enhanced CO<sub>2</sub> uptake by the oceans and a decrease in

**Competing interests:** The authors have declared that no competing interests exist.

ocean surface water pH, referred to as ocean acidification (OA) [1,2]. From 1765 until 1994, pH values were calculated to have already decreased by 0.08 units. Present-day CO<sub>2</sub> concentrations of around 400 µatm are predicted to more than double by the year 2100, which will result in a further acidification of the ocean [3]. After the Polar Oceans, the North Atlantic is expected to show strongest changes in response to rising pCO<sub>2</sub> [3,4]. As a major sink of anthropogenic CO<sub>2</sub>, the North Atlantic Ocean basin stores almost a quarter of the global oceanic anthropogenic CO<sub>2</sub>, although covering only 15% of the global ocean area [5]. The projected changes in ocean carbonate chemistry may thus not only have strong effects on the marine biota, but also on the oceanic carbon cycling.

Phytoplankton take up inorganic carbon (C<sub>i</sub>) in the photic zone and fix it into organic compounds, thereby providing a carbon and energy source for higher trophic levels. The key enzyme of carbon fixation, the CO<sub>2</sub>-binding enzyme Ribulose 1,5-bisphosphate Carboxylase/Oxygenase (RubisCO), exhibits a generally low affinity for its substrate CO<sub>2</sub> [6–8]. To avoid C<sub>i</sub> limitation, many phytoplankton species operate carbon concentrating mechanisms (CCMs) [9,10]. The efficiency in CO<sub>2</sub> fixation depends on both the type of RubisCO as well as the mode of CCMs so that the response of phytoplankton to OA cannot be generalized across taxa [11–13]. Various studies have provided mechanistic insights into the CO<sub>2</sub>-dependent regulation of CCMs and thus CO<sub>2</sub> fixation over a range of phytoplankton species (e.g. [14–16]). Besides species-specific differences, also strains of the same species may respond differently (e.g. [17–19]), which further complicates predictions on OA-driven changes in primary production.

To test these effects directly, numerous studies have exposed natural phytoplankton communities to high pCO<sub>2</sub>, either in bottle incubations or mesocosms, often finding higher rates of CO<sub>2</sub> fixation under OA [20]. In these experiments, which lasted only a couple of days up to a month, the effects were yet relatively small. Here, we investigated the impact of OA on primary production by a natural phytoplankton community over an entire winter-to-summer succession. Experiments were performed in large scale mesocosms, deployed in the Gullmar Fjord located in Southwest Sweden at the Skagerrak coast in 2013 [21]. Depending on the wind direction and tides, the fjord consists of high saline bottom water from the North Atlantic, a low salinity thin surface layer fed with water from the river Örekil, and in between a layer fed by the Baltic current. Monitoring data from over 100 years have shown that the phytoplankton spring community in the Gullmar Fjord is typically dominated by diatoms, whereas summer blooms often comprise dinoflagellates [22,23]. We assessed primary production of the phytoplankton community from the mesocosms as well as the fjord by applying <sup>14</sup>C incubations over 24 h [24]. We furthermore assessed the light dependency of CO<sub>2</sub> fixation by performing photosynthesis-irradiance response curves in short incubations (80 min.).

## Material and methods

### Primary production experiments did not involve endangered or protected species

The KOSMOS 2013 mesocosm experiment was performed in the Gullmar Fjord (Kristineberg, Sweden) from March until July 2013 as part of the project BIOACID (Biological Impacts of Ocean ACIDification) phase II. Ten mesocosms were deployed near Kristineberg, with permission from the Sven Lovén Centre for Marine Infrastructure. The mesocosms were cylindrical polyurethane bags with a 2 m diameter mounted in a floatation frame [25]. The bags reached a depth of 17 m and were closed at the bottom with a 2 m long conical sediment trap [26]. Two days prior to the experiment (i.e. t-2), a water body was enclosed inside the mesocosms by lifting the upper end about one meter above the surface.

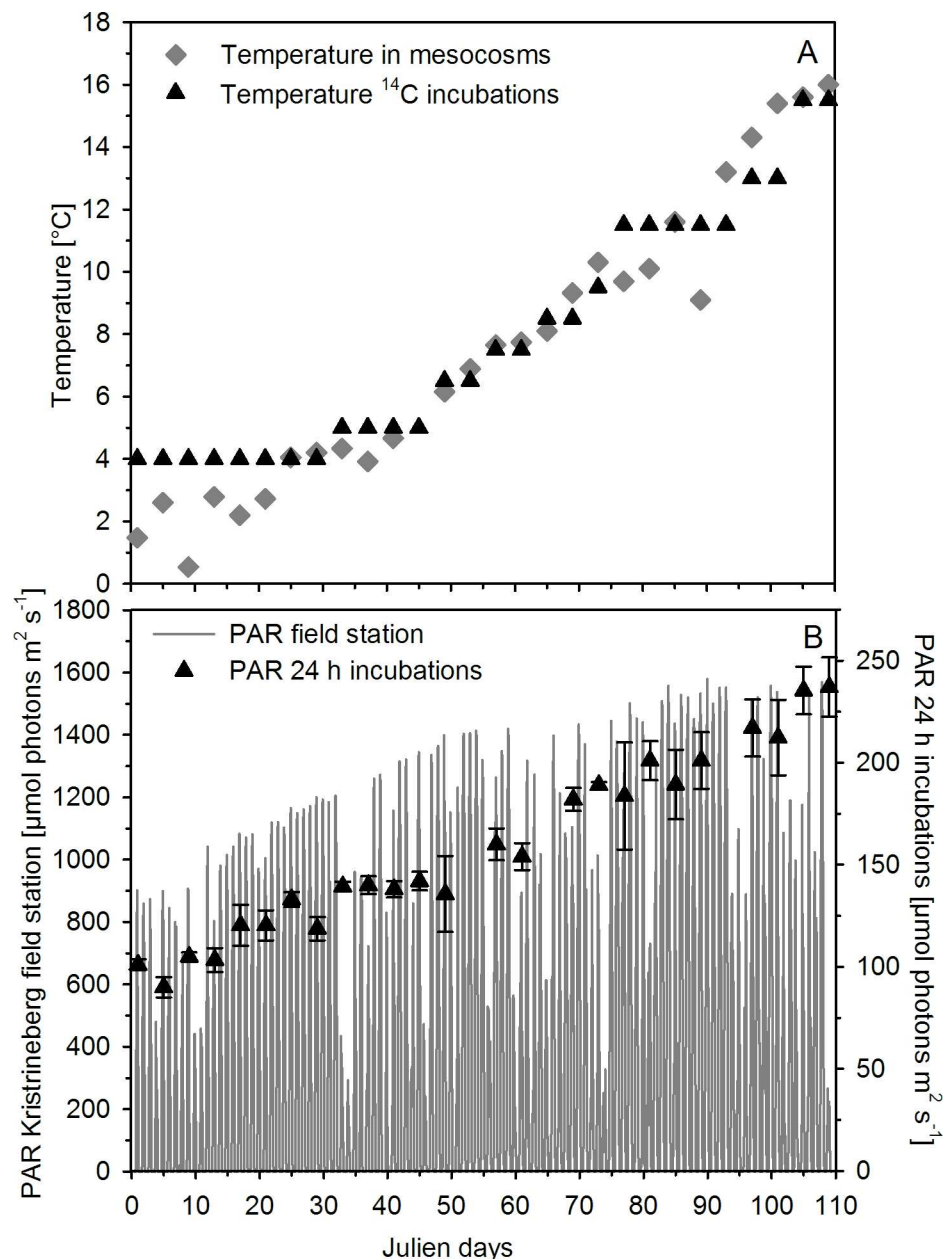
All mesocosms had a salinity of about 29, and nitrate, phosphate and silicate concentrations of about 7, 0.8, and 10  $\mu\text{mol L}^{-1}$ , respectively.  $\text{CO}_2$  enrichment was conducted on t-1 and t0, for which sterile-filtered and  $\text{CO}_2$ -saturated seawater from the Gullmar Fjord was added to five mesocosms (M2, M4, M6, M7, M8). The remaining five mesocosms (M1, M3, M5, M9, M10) were treated as controls and received no  $\text{CO}_2$ -enriched seawater. Average  $p\text{CO}_2$  (based on dissolved inorganic carbon (DIC) and spectrophotometric  $\text{pH}_T$  measurements) in the 'low' and 'high'  $\text{CO}_2$  treatments were about 380 and 760  $\mu\text{atm}$ , respectively. The systems were open and allowed a gas exchange at the sea surface. To account for  $\text{CO}_2$  losses to the atmosphere by outgassing and for  $\text{CO}_2$  consumption by primary production,  $\text{CO}_2$  was added on a regular basis to the 'high'  $\text{CO}_2$  treatments. As a consequence,  $\text{CO}_2$  concentrations remained above the control treatment at all times (for more details see [21]). Sampling of seawater from each mesocosm was done with a depth-integrated water sampler (Hydro-Bios). After initial sampling on t0 and t1, samples were taken every other day until t109 (i.e. t3, t5, t7 etc.). For further information on the design and set-up of the experiment, as well as the  $\text{CO}_2$  perturbation and sampling techniques, we refer to [21].

## Sampling for primary production

For our measurements, integrated water samples from 0–17 m depth were taken in a four day interval (i.e. t1, t5, t9, etc.) from each of the ten mesocosms, and an additional sample was taken from the fjord. Sampling usually took place between 9 and 12 a.m. and aliquots from well mixed water samples were filled in gas-tight and headspace-free bottles (Schott) of 250 mL (for the 24 h incubations) and 500 mL (for the photosynthesis-irradiance response (P/I) curves). Samples were brought directly to the laboratory, where they were gently filtered over a 500  $\mu\text{m}$  mesh-size filter to remove larger zooplankton from the samples, and were kept at the *in situ* water temperature until incubations started. Over the course of the entire experiment, the temperature in the fjord increased from 1.5°C at t1 towards 15.5°C at t109, and we adjusted the incubation temperatures accordingly (Fig 1A). Only at the beginning of the experiment, when productivity and biomass was still low, we could not fully match the temperature from the fjord as our incubator was not able to maintain temperatures below 4°C. Light was provided by daylight tubes (OSRAM) from the side in a 16:8 h light-dark cycle. To account for the increase in light intensities over the course of the experiment in the mesocosms, the light intensity was stepwise increased in the incubator (Fig 1B). Using a spherical micro quantum sensor (Walz), we increased the photon flux density (PFD) every 16 days (i.e. after 4 sampling days) by about 20  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ , starting with around 100  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$  at t1 and ending with 240  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$  at t109.

## Primary production measurements

Primary productivity was measured according to Steeman Nielsen [24]. Despite limitations [27], this approach has remained the method of choice, especially for field work, as it allows assessing rates even at times of low productivity. One has to keep in mind, however, that measured rates have different meanings depending on the incubation time [27]. In our 80 min. incubations for  $^{14}\text{C}$ -based photosynthesis-irradiance response (P/I) curves, we obtained rates of gross primary production because there is only little loss of incorporated  $^{14}\text{C}$  via respiration and exudation over such short timescales. In our 24 h incubations for the  $^{14}\text{C}$ -based primary production measurements, respiration lowers the  $^{14}\text{C}$  incorporation and thus net rates of primary production are obtained. To account for fixed  $^{14}\text{C}$  ending up in the dissolved phase, which can be a significant proportion under nutrient deplete conditions, we included values of the filtrate in our  $\text{PP}_T$  estimates.



**Fig 1.** Mean temperature in mesocosms (grey diamonds) and during <sup>14</sup>C incubations (black triangles) (A), and incoming light (PAR) at the Kristineberg field station around midday (<http://www.weather.loven.gu.se/kristineberg/en>; grey lines) and during <sup>14</sup>C-based 24 h incubations (black triangles) (B). Triangles indicate the mean  $\pm$  SD of three light measurements from the bottom, middle and top of a representative incubation vial.

doi:10.1371/journal.pone.0172594.g001

### <sup>14</sup>C-based primary production measurements

For the 24 h incubations, 40 mL sample volumes were spiked with 20  $\mu\text{L}$  of <sup>14</sup>C-labeled sodium bicarbonate ( $\text{NaH}^{14}\text{CO}_3$ ; from a  $1 \text{ mCi mL}^{-1} = 37 \text{ MBq mL}^{-1}$  stock solution; PerkinElmer). Two incubation vials for each mesocosm, and the fjord water, were prepared accordingly (i.e. 22 vials in total, of which 11 were used for the light and the remaining 11 for the dark incubations). Determination of total <sup>14</sup>C-spike addition was done from an

extra 40 mL  $^{14}\text{C}$ -spiked water sample. For this purpose, 1 mL was directly transferred into a 20 mL scintillation vial (PerkinElmer) containing 10 mL scintillation cocktail (Ultima Gold AB; PerkinElmer) and counted in a liquid scintillation analyzer (Beckman LS6500). Blank determination was done by transferring 1 mL from the extra  $^{14}\text{C}$ -spiked water sample into 6 mL of 6 M HCl, which degassed for 48 h and was then counted after adding 10 mL scintillation cocktail. All incubations were placed on an orbital shaker in a temperature-controlled incubator.

Incubations were stopped after 24 h by vacuum filtration onto GF/F filters (Whatman). To estimate the amount of  $\text{C}_i$  fixation into particulate organic carbon (POC), filters were rinsed twice with 20 mL of sterile filtered seawater (0.2  $\mu\text{m}$ ), and subsequently placed in scintillation vials containing 300  $\mu\text{L}$  of 3 M HCl to remove  $^{14}\text{C}$ -labeled DIC. To estimate the amount of  $\text{C}_i$  fixation ending up in the pool of dissolved organic carbon (DOC), 6 mL of filtrate was transferred into a scintillation vial, acidified with 1 mL 6 M HCl, and placed under a fume hood for 48 hours for degassing  $\text{D}^{14}\text{C}$ . Prior to measurements, 10 mL of scintillation cocktail was added to each vial and filter, thoroughly mixed, and counted in a liquid scintillation analyzer. Primary production (PP) was calculated according to:

$$PP = \frac{DIC \cdot (DPM_{\text{sample}} - DPM_{\text{blank}}) \cdot 1.05}{(DPM_{100\%} \cdot t)} \quad \text{Eq. 1}$$

where  $DPM$  represents the decays per minute and  $t$  represents time. Correction for non-specific  $^{14}\text{C}$  fixation in the dark was done by subtracting dark incubations from light incubations. Dark  $^{14}\text{C}$  fixation accounted for about 1 to 6% of the light incubations during times of high and low productivity, respectively. Based on the phytoplankton community composition [21], some primary producers were smaller than the pore size of our filters (i.e.  $<0.7 \mu\text{m}$ ). We therefore reported total primary production ( $\text{PP}_T$ ;  $\mu\text{mol C L}^{-1} \text{h}^{-1}$ ) from the 24 h incubations as the sum of  $\text{CO}_2$  fixation into POC and DOC.

### $^{14}\text{C}$ -based photosynthesis-irradiance response curves

For the photosynthesis-irradiance response (P/I) curves, 300 mL sample volume from each mesocosm was spiked with 100  $\mu\text{Ci}$  of  $\text{NaH}^{14}\text{CO}_3^-$  (PerkinElmer) and subdivided into seven 40 mL glass vials. From the remaining  $^{14}\text{C}$ -spiked seawater, 200  $\mu\text{L}$  aliquots were transferred into a 10 mL scintillation cocktail to determine total spike addition for each P/I curve. While one vial was incubated in the dark, the six remaining vials were exposed to increased light intensities ranging from about 10 to 700  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$  in a custom-made photosynthetron. Light was supplied from below and the PFD was assessed prior to each experimental day. The photosynthetron was placed in the same incubator as the 24 h incubations. Additional temperature control was achieved via a water bath connected to the sample holder. After an incubation time of 80 min. at the respective light conditions, samples were filtered on GF/F filters (Whatman). Analysis of  $\text{PO}^{14}\text{C}$  was determined following the same procedure as for the 24 h incubations and data was fitted according to:

$$PP_{P/I} = P_{\text{max}} \cdot (1 - e^{-\alpha \cdot (I - I_k)}) \quad \text{Eq. 2}$$

where  $P_{\text{max}}$  is the light-saturated rate of photosynthesis,  $\alpha$  is the light-limited (i.e. initial) slope of the P/I curve representing the maximum light-use efficiency,  $I$  is the irradiance, and  $I_k$  is the light saturation index. Rates of  $\text{PP}_{P/I}$  were normalized to chlorophyll  $a$  (Chl  $a$ ) concentrations in the samples from the particular day and mesocosm [21].

## Statistics

Differences in  $PP_T$ , Chl *a*,  $P_{max}$ ,  $I_k$  and  $\alpha$  between the  $CO_2$  treatments were tested over time by a two-way repeated measures Analysis of Variance (rmANOVA), and the association between  $PP_T$  and Chl *a* was tested by Pearson product-moment correlations. Variables were log+1 or square root transformed if this improved normality or homogeneity of variances, as tested by the Shapiro-Wilk test or Levene's test, respectively. All statistics were performed with SigmaPlot 12.5 (Systat).

## Results

### Total primary production

For the first three weeks of the experiment, estimates on  $PP_T$  were lower in the mesocosms than in the fjord (Fig 2A). All mesocosms showed comparable development in  $PP_T$ , with an initial period of low productivity (phase I, t1-t16), a first spring bloom of highest productivity around t33 (phase II, t17-t40), followed by a second bloom of highest productivity around t57 (phase III, t41-77), and a subsequent period of low productivity until the end of the experiment (phase IV, t78-t109; Fig 2A, Table 1). Dynamics in primary production in the mesocosms differed from that in the fjord. For example,  $PP_T$  was higher in the fjord during phase I, while  $PP_T$  was higher in the mesocosms during phase II. Also, a small increase in  $PP_T$  present in the fjord at the start of phase IV was lacking in the mesocosms (Fig 2A).

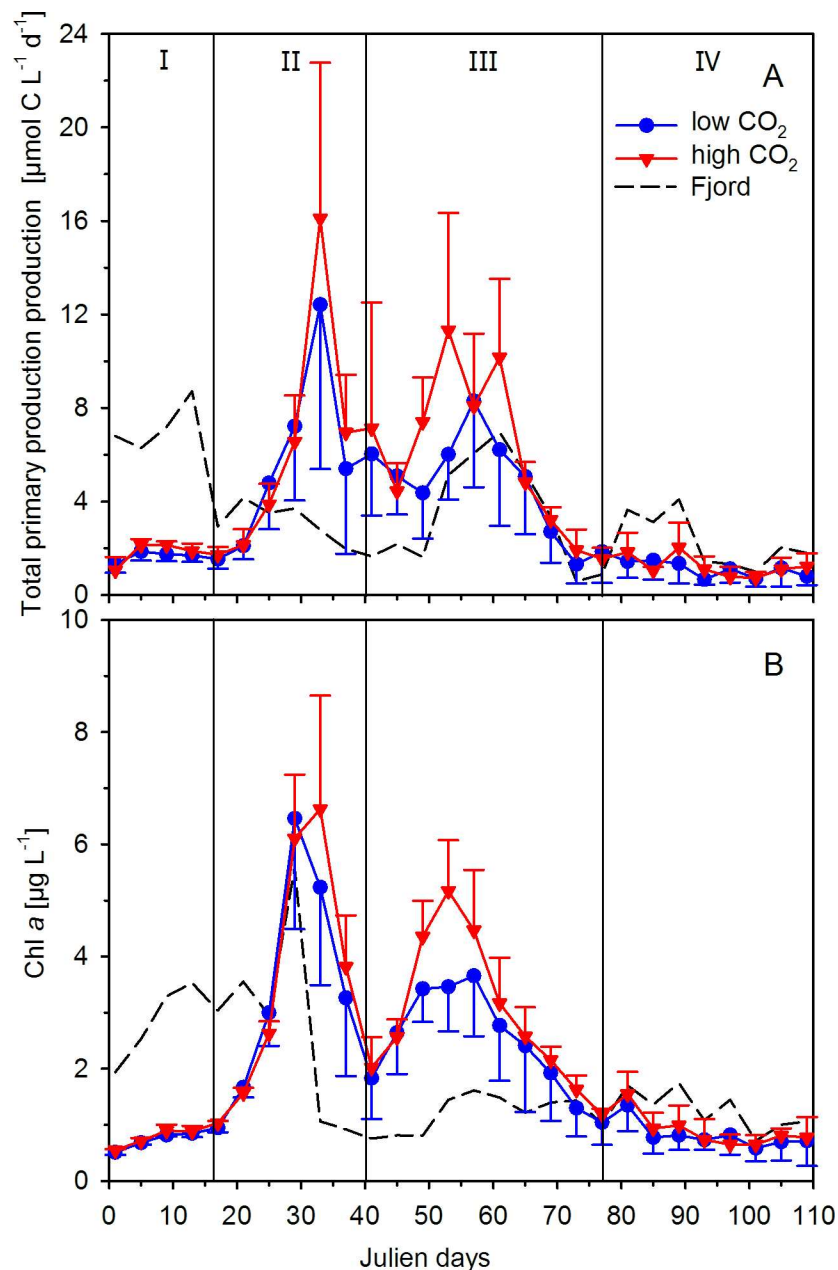
High  $pCO_2$  yielded higher mean estimates on  $PP_T$  during both blooms, although differences during both blooms were not significant (Table 2). Highest  $PP_T$  was observed during the first bloom at t33, with up to  $16.1 \pm 6.7 \mu\text{mol C L}^{-1} \text{ d}^{-1}$  at high  $pCO_2$  and  $12.4 \pm 7.0 \mu\text{mol C L}^{-1} \text{ d}^{-1}$  at low  $pCO_2$ . During the second bloom,  $PP_T$  amounted to highest values of  $11.3 \pm 5.0 \mu\text{mol C L}^{-1} \text{ d}^{-1}$  at t53 for high  $pCO_2$ , and  $6.0 \pm 1.9 \mu\text{mol C L}^{-1} \text{ d}^{-1}$  at day t57 for low  $pCO_2$ . At the peak of the second bloom,  $PP_T$  appeared to be higher at high  $pCO_2$ , though this difference was marginally significant and dependent on time (Table 2; rmANOVA, Time x  $CO_2$  treatment,  $P = 0.098$ ). During both blooms phases, Chl *a* remained unaltered in response to OA (Table 2), though at times showed higher concentrations at high  $pCO_2$  [21]. Furthermore, Chl *a* was strongly correlated to  $PP_T$  ( $\sigma = 0.87$ ,  $P < 0.0001$ ).

When cumulated over the experimental period of 109 days, the  $PP_T$  data yielded a total of  $92 \pm 29.21$  and  $110 \pm 25.79 \mu\text{mol C L}^{-1}$  at low and high  $pCO_2$ , respectively. In the fjord, cumulative  $PP_T$  yielded  $95 \mu\text{mol C L}^{-1}$  (Fig 3A), being more comparable to  $PP_T$  in the mesocosms at low  $pCO_2$ . The difference in cumulative  $PP_T$  between low and high  $pCO_2$  was about 20% and closely matched the observed difference in Chl *a* concentration of about 15%. Consequently, no differences in the yields were observed when normalizing cumulated  $PP_T$  to Chl *a* (as to account for changes in phytoplankton biomass). In both treatments, we got a cumulative value of around  $600 \mu\text{g C } (\mu\text{g Chl } a)^{-1}$  until the end of the experiment (Fig 3B). Chl *a*-normalized cumulative  $PP_T$  in the fjord was higher than in the mesocosms and amounted to a total of about  $700 \mu\text{g C } (\mu\text{g Chl } a)^{-1}$  (Fig 3B).

### Photoacclimation

P/I curves provided information on the photoacclimation of the phytoplankton communities in the mesocosms and the fjord.  $P_{max}$  was on average  $3.17 \pm 0.54$  and  $3.38 \pm 0.26 \mu\text{g C } (\mu\text{g Chl } a)^{-1} \text{ h}^{-1}$  at low and high  $pCO_2$ , respectively. There was no apparent  $CO_2$  effect on  $P_{max}$  during both blooms (Table 2), which furthermore strongly varied between mesocosms and sampling days (Fig 4A).  $I_k$ , indicating the light intensity at which phytoplankton shifts from light limitation to light saturation, changed over the course of the experiment (Fig 4B). More specifically,





**Fig 2.** Mean values of total primary production (from  $^{14}\text{C}$ -based 24 h incubations; A) and chlorophyll a concentrations (B) from mesocosm and fjord samples. Triangles (red; high  $p\text{CO}_2$ ) and circles (blue; low  $p\text{CO}_2$ ) represent the mean  $\pm$  SD of five biological replicates. Roman numbers denote the different phases of the experiment.

doi:10.1371/journal.pone.0172594.g002

in the period prior to the first bloom (phase I),  $I_k$  remained around  $100 \mu\text{mol photons m}^{-2} \text{s}^{-1}$  and increased towards the end of the first bloom phase reaching mean values of approximately 160 and  $250 \mu\text{mol photons m}^{-2} \text{s}^{-1}$  at high and low  $p\text{CO}_2$ , respectively. In the course of the second bloom,  $I_k$  decreased resulting in lowest values of  $50 \mu\text{mol photons m}^{-2} \text{s}^{-1}$  around t61 (Fig 4), after which it increased again to values of around  $150 \mu\text{mol photons m}^{-2} \text{s}^{-1}$  (Fig 4B). Besides these general changes over the season, we did not observe a significant  $\text{CO}_2$  effect on  $I_k$  values during both blooms (Table 2). The maximum light-use efficiency also changed in the course of

**Table 1. Total primary production ( $\mu\text{mol C L}^{-1} \text{d}^{-1}$ ) in the mesocosms derived from 24 h incubations.** Values at high  $p\text{CO}_2$  are indicated in bold letters (M2, M4, M6–8). Grey shading indicates the peak of the two bloom phases.

Julien day	M1	M2	M3	M4	M5	M6	M7	M8	M9	M10
1	1.88	<b>1.34</b>	0.85	<b>1.42</b>	1.44	<b>1.46</b>	<b>0.95</b>	<b>0.08</b>	1.16	1.28
5	1.75	<b>1.97</b>	1.29	<b>2.04</b>	2.15	<b>2.58</b>	<b>2.12</b>	<b>1.96</b>	1.82	2.26
9	1.96	<b>2.16</b>	1.25	<b>2.30</b>	1.93	<b>2.27</b>	<b>1.96</b>	<b>2.01</b>	1.93	1.65
13	1.69	<b>1.60</b>	1.26	<b>2.22</b>	2.02	<b>2.22</b>	<b>1.81</b>	<b>1.57</b>	1.80	1.68
17	1.48	<b>1.53</b>	1.83	<b>1.48</b>	0.85	<b>1.43</b>	<b>2.13</b>	<b>2.03</b>	1.88	1.61
21	2.63	<b>2.15</b>	2.04	<b>2.46</b>	1.77	<b>3.07</b>	<b>1.34</b>	<b>1.66</b>	1.34	2.72
25	6.08	<b>4.60</b>	2.81	<b>4.22</b>	2.65	<b>2.92</b>	<b>4.70</b>	<b>2.98</b>	5.34	7.05
29	9.48	<b>6.88</b>	4.68	<b>8.39</b>	3.27	<b>6.45</b>	<b>7.77</b>	<b>3.30</b>	7.88	10.79
33	15.94	<b>5.04</b>	4.92	<b>22.40</b>	13.20	<b>15.47</b>	<b>18.89</b>	<b>18.73</b>	6.16	21.83
37	7.70	<b>3.40</b>	3.29	<b>9.70</b>	4.05	<b>7.62</b>	<b>8.37</b>	<b>5.65</b>	1.45	10.47
41	8.91	<b>3.08</b>	5.25	<b>16.59</b>	2.02	<b>5.11</b>	<b>5.17</b>	<b>5.68</b>	6.32	7.65
45	5.10	<b>2.73</b>	6.98	<b>5.25</b>	2.50	<b>5.48</b>	<b>5.07</b>	<b>3.66</b>	5.14	5.70
49	5.65	<b>8.65</b>	2.45	<b>8.10</b>	3.43	<b>8.60</b>	<b>4.10</b>	<b>7.59</b>	3.16	7.15
53	9.01	<b>4.90</b>	4.93	<b>14.08</b>	5.80	<b>12.51</b>	<b>17.43</b>	<b>7.65</b>	3.89	6.41
57	9.69	<b>6.15</b>	3.91	<b>9.96</b>	9.95	<b>10.87</b>	<b>9.84</b>	<b>3.59</b>	12.80	5.10
61	6.77	<b>7.60</b>	2.78	<b>13.59</b>	3.42	<b>6.64</b>	<b>13.83</b>	<b>9.15</b>	7.24	10.85
65	5.77	<b>4.71</b>	1.24	<b>4.10</b>	4.49	<b>6.21</b>	<b>4.98</b>	<b>4.08</b>	5.90	7.90
69	3.13	<b>2.54</b>	1.06	<b>3.80</b>	1.85	<b>2.83</b>	<b>3.69</b>	<b>3.17</b>	4.55	2.97
73	1.05	<b>1.75</b>	0.69	<b>1.71</b>	0.65	<b>1.84</b>	<b>3.36</b>	<b>0.92</b>	1.58	2.62
77	3.68	<b>0.90</b>	0.85	<b>1.94</b>	0.53	<b>1.97</b>	<b>1.70</b>	<b>1.22</b>	1.41	2.77
81	2.40	<b>0.39</b>	0.72	<b>2.05</b>	0.86	<b>1.93</b>	<b>2.41</b>	<b>2.40</b>	1.36	1.79
85	2.16	<b>1.28</b>	1.35	<b>0.88</b>	0.75	<b>0.98</b>	<b>1.07</b>	<b>1.01</b>	0.65	2.49
89	1.10	<b>1.13</b>	1.02	<b>1.52</b>	1.08	<b>1.40</b>	<b>3.74</b>	<b>2.37</b>	0.67	2.83
93	0.29	<b>0.52</b>	0.73	<b>1.23</b>	0.81	<b>0.80</b>	<b>1.98</b>	<b>0.94</b>	0.83	0.65
97	1.29	<b>0.57</b>	1.01	<b>0.43</b>	0.19	<b>0.47</b>	<b>1.44</b>	<b>0.98</b>	1.27	1.79
101	0.83	<b>0.63</b>	0.41	<b>1.02</b>	1.30	<b>0.64</b>	<b>0.44</b>	<b>0.95</b>	0.50	0.52
105	0.78	<b>0.85</b>	1.91	<b>0.99</b>	2.11	<b>1.98</b>	<b>0.86</b>	<b>0.85</b>	0.35	0.62
109	1.05	<b>1.65</b>	0.48	<b>0.65</b>	1.34	<b>0.54</b>	<b>1.63</b>	<b>1.60</b>	0.48	0.66

doi:10.1371/journal.pone.0172594.t001

the phytoplankton succession. Highest  $\alpha$  values coincided with the phytoplankton blooms during phases II and III and were observed around t30 and t56 in all mesocosms (Fig 4C). Similar to the other parameters, there was no significant  $\text{CO}_2$  effect on  $\alpha$  values during both blooms (Table 2).

## Discussion

We did not observe a sustained effect of OA on primary production during the investigated winter-to-summer plankton succession. When focusing on the peak of the second spring bloom in phase III, however,  $\text{PP}_T$  showed a marginally significant increase under high  $p\text{CO}_2$  (Table 2). During this distinct phase, the availability of inorganic nutrients was low and primary production was fueled by *in situ* remineralization [21]. Integrated over the entire experimental period, OA yielded about 20% more  $\text{CO}_2$  fixation. Such enhanced primary production is in line with the higher Chl *a* concentration under these conditions.

At the onset of the experiment, concentrations of major nutrients in the mesocosms were higher than in the fjord [21]. The lower concentrations in the fjord were the result of higher primary production compared to the mesocosms right after closure of the mesocosms



**Table 2. Output of the repeated measures ANOVA for phase II, phase III, peak of bloom 1 and peak of bloom 2, with degrees of freedom (df), the *F*-value and the *P*-value.** Significant outcomes are indicated with  $P < 0.001$  (\*\*\*),  $P < 0.01$  (\*\*),  $P < 0.05$  (\*) and  $P < 0.1$  (·).

	Parameter	Effect	df	<i>F</i>	<i>P</i>
Phase II (incl. bloom 1; t17-t40)	PP <sub>T</sub> (μmol C L <sup>-1</sup> d <sup>-1</sup> )	CO <sub>2</sub> treatment	1	0.278	0.612
		Time	5	38.060	<0.001***
		Time x CO <sub>2</sub> treatment	5	0.814	0.547
	Chl <i>a</i> (μg Chl <i>a</i> L <sup>-1</sup> )	CO <sub>2</sub> treatment	1	0.228	0.646
		Time	5	13.818	<0.001***
		Time x CO <sub>2</sub> treatment	5	0.245	0.940
	P <sub>max</sub> (μg C (μg Chl <i>a</i> ) <sup>-1</sup> h <sup>-1</sup> )	CO <sub>2</sub> treatment	1	0.845	0.383
		Time	5	15.796	<0.001***
		Time x CO <sub>2</sub> treatment	5	0.497	0.776
	<i>I<sub>K</sub></i> (μmol photons m <sup>-2</sup> s <sup>-1</sup> )	CO <sub>2</sub> treatment	1	0.651	0.443
		Time	5	2.647	0.037*
		Time x CO <sub>2</sub> treatment	5	0.712	0.618
Phase III (incl. bloom 2; t41-t77)	PP <sub>T</sub> (μmol C L <sup>-1</sup> d <sup>-1</sup> )	CO <sub>2</sub> treatment	1	1.481	0.258
		Time	9	26.124	<0.001***
		Time x CO <sub>2</sub> treatment	9	1.566	0.142
	Chl <i>a</i> (μg Chl <i>a</i> L <sup>-1</sup> )	CO <sub>2</sub> treatment	1	0.395	0.547
		Time	9	9.258	<0.001***
		Time x CO <sub>2</sub> treatment	9	0.915	0.517
	P <sub>max</sub> (μg C (μg Chl <i>a</i> ) <sup>-1</sup> h <sup>-1</sup> )	CO <sub>2</sub> treatment	1	1.538	0.250
		Time	9	4.126	<0.001***
		Time x CO <sub>2</sub> treatment	9	0.565	0.821
	<i>I<sub>K</sub></i> (μmol photons m <sup>-2</sup> s <sup>-1</sup> )	CO <sub>2</sub> treatment	1	0.181	0.681
		Time	9	4.544	<0.001***
		Time x CO <sub>2</sub> treatment	9	0.422	0.919
Peak of bloom 1 (t29-t33)	PP <sub>T</sub> (μmol C L <sup>-1</sup> d <sup>-1</sup> )	CO <sub>2</sub> treatment	1	0.340	0.576
		Time	1	13.682	0.006**
		Time x CO <sub>2</sub> treatment	1	1.194	0.306
	Chl <i>a</i> (μg Chl <i>a</i> L <sup>-1</sup> )	CO <sub>2</sub> treatment	1	0.092	0.769
		Time	1	50.114	<0.001***
		Time x CO <sub>2</sub> treatment	1	0.152	0.707
	P <sub>max</sub> (μg C (μg Chl <i>a</i> ) <sup>-1</sup> h <sup>-1</sup> )	CO <sub>2</sub> treatment	1	0.372	0.559
		Time	1	38.768	<0.001***
		Time x CO <sub>2</sub> treatment	1	0.017	0.899
	<i>I<sub>K</sub></i> (μmol photons m <sup>-2</sup> s <sup>-1</sup> )	CO <sub>2</sub> treatment	1	0.858	0.381
		Time	1	0.590	0.465
		Time x CO <sub>2</sub> treatment	1	0.581	0.468
	Alpha	CO <sub>2</sub> treatment	1	0.360	0.565
		Time	1	33.248	<0.001***
		Time x CO <sub>2</sub> treatment	1	0.653	0.442

(Continued)

Table 2. (Continued)

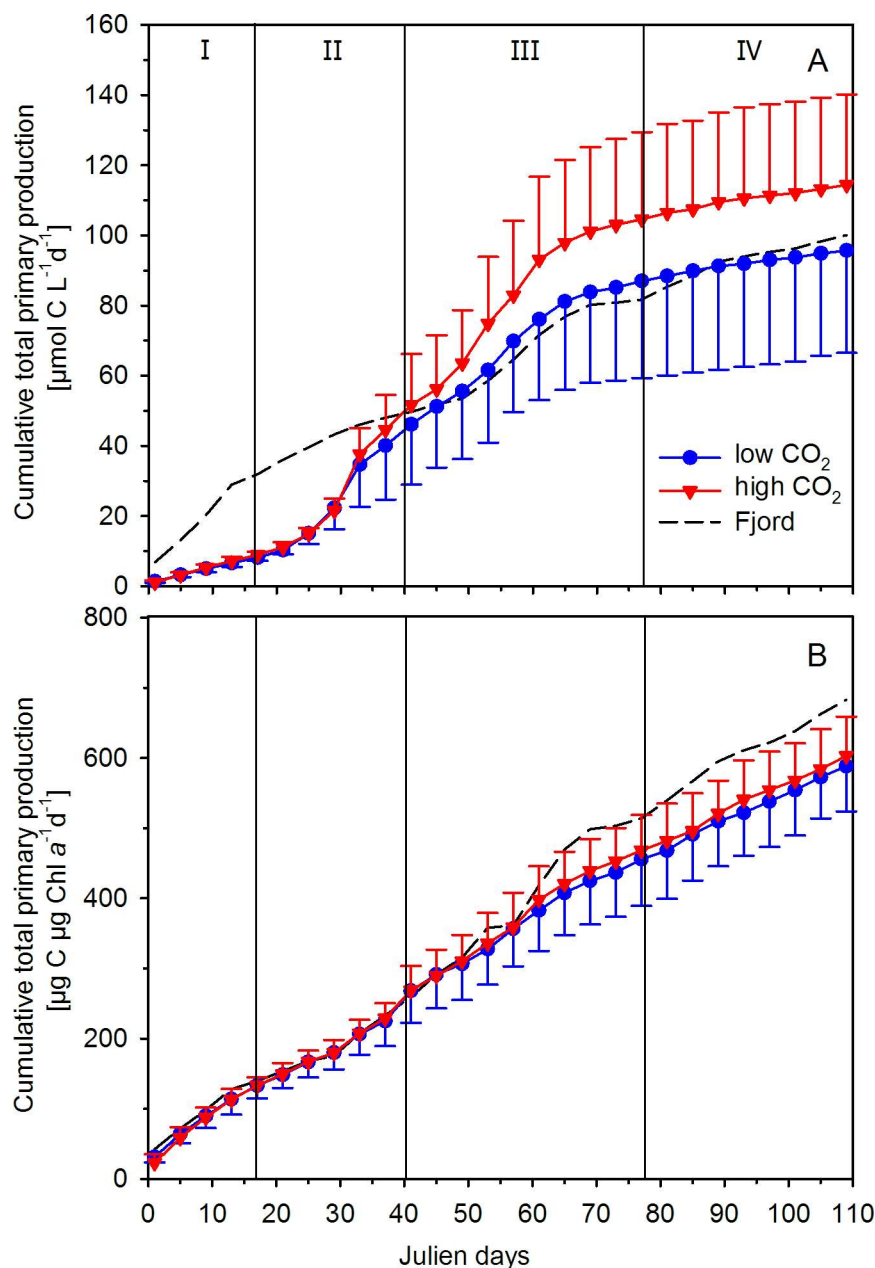
	Parameter	Effect	df	F	P
Peak of bloom 2 (t53-t61)	PP <sub>T</sub> (μmol C L <sup>-1</sup> d <sup>-1</sup> )	CO <sub>2</sub> treatment	1	3.134	0.115
		Time	2	0.099	0.907
		Time x CO <sub>2</sub> treatment	2	2.701	0.098
	Chl <i>a</i> (μg Chl <i>a</i> L <sup>-1</sup> )	CO <sub>2</sub> treatment	1	1.200	0.305
		Time	2	2.168	0.147
		Time x CO <sub>2</sub> treatment	2	2.278	0.135
	P <sub>max</sub> (μg C (μg Chl <i>a</i> ) <sup>-1</sup> h <sup>-1</sup> )	CO <sub>2</sub> treatment	1	0.506	0.497
		Time	2	9.666	0.002**
		Time x CO <sub>2</sub> treatment	2	0.029	0.972
	<i>I<sub>k</sub></i> (μmol photons m <sup>-2</sup> s <sup>-1</sup> )	CO <sub>2</sub> treatment	1	0.120	0.738
		Time	2	5.210	0.018**
		Time x CO <sub>2</sub> treatment	2	1.874	0.186
	Alpha	CO <sub>2</sub> treatment	1	0.072	0.795
		Time	2	1.366	0.283
		Time x CO <sub>2</sub> treatment	2	0.311	0.737

doi:10.1371/journal.pone.0172594.t002

(beginning at t-2, Fig 2). Although initial conditions in the mesocosms were largely comparable to the situation in the fjord, perturbations induced during the set-up of the mesocosms [21], e.g. the water column mixing (t0) or the establishment of CO<sub>2</sub> treatments (t-1 and t0), may have contributed to the delay in primary production.

In the mesocosms, PP<sub>T</sub> as well as Chl *a* concentrations remained relatively low during phase I and started to increase more pronounced around t20, leading to a first phytoplankton bloom with highest PP<sub>T</sub> around t33 (Fig 2). During this phase II, major nutrients such as inorganic phosphate and nitrogen were depleted to very low values (for more details, see Bach et al. [21]). This nutrient depletion, particularly for nitrogen, together with grazing presumably caused the collapse of the phytoplankton bloom and the decrease in PP<sub>T</sub> as well as Chl *a* concentrations (Fig 2). At the same time at the Kristineberg field station (~3 km distance to mesocosm deployment site), a sudden drop in the *in situ* light intensity was detected (Fig 1B), coinciding with the peak of the first bloom (Fig 2A). In fact, the average water column light intensities (0–19 m depth) during midday for all ten mesocosms were reduced to about 35 μmol photons m<sup>-2</sup> s<sup>-1</sup> for several days (Fig 4B), and dropped below the *I<sub>k</sub>* values (about 115 μmol photons m<sup>-2</sup> s<sup>-1</sup>). Such lower light levels may possibly have, at least temporally, limited photosynthesis and thereby affected the response of phytoplankton to low nutrient levels. While there were dynamic changes in *P<sub>max</sub>*, *I<sub>k</sub>*, and *α* over the course of the phytoplankton succession (Fig 4), there was no effect of OA on photoacclimation.

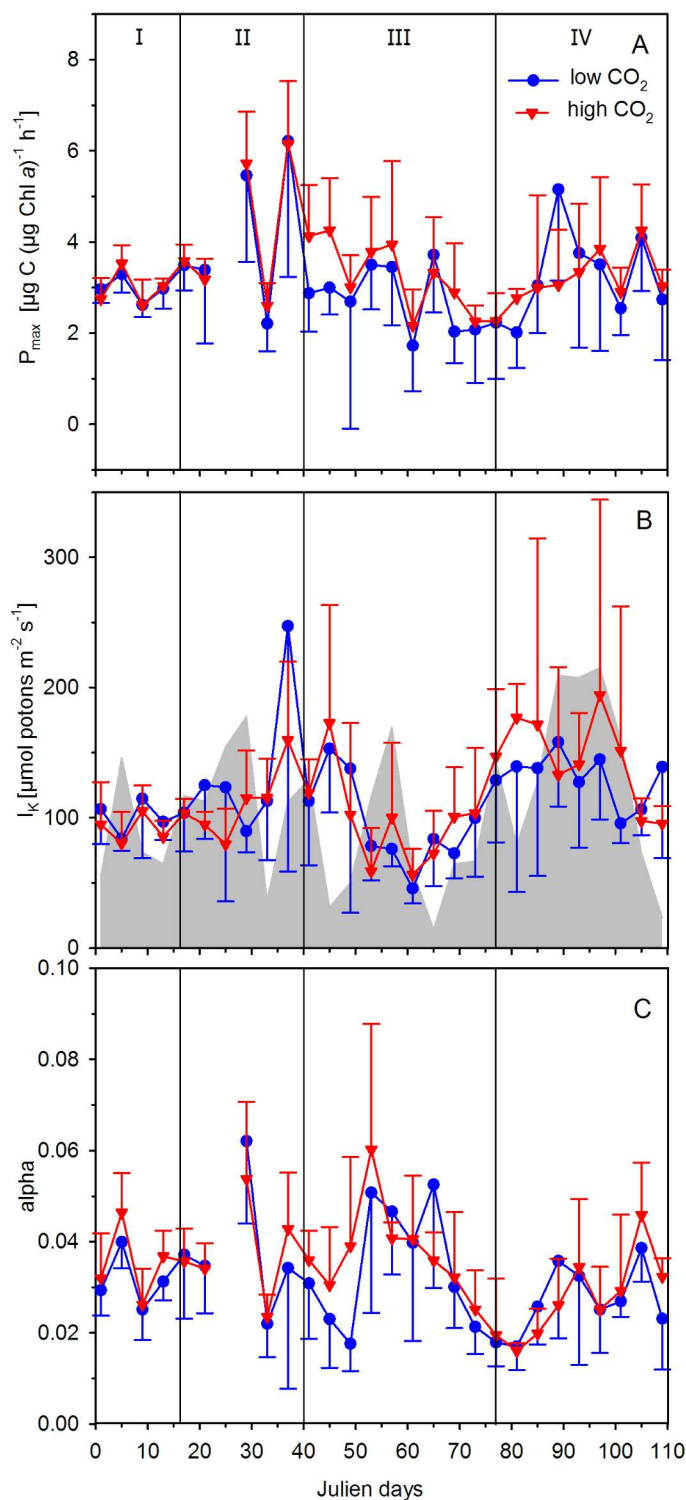
Dissolved phosphate and inorganic nitrogen concentrations remained low during phase III, while PP<sub>T</sub> and Chl *a* concentrations increased again, causing the second bloom (Fig 2). An earlier study in the Gullmar Fjord also showed a relatively high primary production during summer months, despite low nutrient concentrations [28,29]. According to this long-term Gullmar Fjord time-series study, nutrients were not only derived from recycled production, but also from local precipitation, run-off, and input from the Kattegat [29]. As the mesocosms were isolated from the surrounding water, nutrient input for primary production should have derived from regeneration only. In fact, dissolved organic nitrogen and NH<sub>4</sub><sup>+</sup> concentrations in the mesocosms remained low, indicating a rapid cycling of nutrients in the food web [21]. Interestingly, it is under these conditions of recycled production and low concentrations of inorganic nutrients that we observed the strongest response in PP<sub>T</sub> towards OA (Fig 2A).



**Fig 3.** Cumulative total primary production (from  $^{14}\text{C}$ -based 24 h incubations; A) and normalized to chlorophyll *a* concentrations (B) from mesocosm and fjord samples. Triangles (red; high  $p\text{CO}_2$ ) and circles (blue; low  $p\text{CO}_2$ ) represent the mean  $\pm$  SD of five biological replicates. Roman numbers denote the different phases of the experiment.

doi:10.1371/journal.pone.0172594.g003

Under nutrient-limited conditions, effects of elevated  $p\text{CO}_2$  on phytoplankton productivity, standing stock and community composition were often found to be stronger [30–32] and comparable findings were also reported with respect to iron limitation [33]. Since nitrogen, phosphorus and iron predominantly limit phytoplankton growth in the global surface oceans [34,35], more studies investigating the combined effects of elevated  $p\text{CO}_2$  and resource limitation are required to provide a mechanistic understanding on the impacts of OA on future primary production.



**Fig 4.** Light-saturated maximum rates ( $P_{max}$ ; A), light saturation index ( $I_k$ ; B), and light-limited slope ( $\alpha$ ; C) of the photosynthesis-response irradiance curves. Triangles (high  $p\text{CO}_2$ ) and circles (low  $p\text{CO}_2$ ) represent the mean  $\pm$  SD of five biological replicates. The grey area in Fig B indicates average water column light intensities (0–19 m depth) during midday for all ten mesocosms. Roman numerals denote the different phases of the experiment.

doi:10.1371/journal.pone.0172594.g004

Even though we did not find a consistent CO<sub>2</sub> response over the entire winter-to-summer plankton succession, the stimulation in primary production under elevated *p*CO<sub>2</sub> at the peak of the second bloom was also observed in earlier studies looking at mixed natural assemblages as well as monoclonal laboratory cultures. During a mesocosm study in Bergen (Norway), for instance, DIC uptake increased under a comparable OA scenario by about 40% [30,36]. Moreover, a mesocosm study in Kongsfjorden (Svalbard, Norway) showed an OA-induced increase in primary production of 10 to 60% over the experimental period [37]. Such increases in primary production may derive from physiological changes in pre-dominant species and/or shifts in community composition both leading to higher phytoplankton biomass buildup. At a higher taxonomic level, the phytoplankton community remained largely unaltered and was dominated by diatoms [21]. Under nutrient-replete as well as nutrient-limiting conditions, elevated *p*CO<sub>2</sub> resulted in an increased abundance of picoeukaryotes [21]. Specific changes within phytoplankton groups will be discussed elsewhere in this special issue (see S1 Table in [21]). With regard to the dominating role of diatoms in our experiment, several studies found this group to enhance their C<sub>i</sub> fixation rates in response to elevated *p*CO<sub>2</sub>, which was often attributed to the down-regulation in the CCM activities under these conditions (e.g. [38–40]). Such enhanced OA-driven efficiencies in C<sub>i</sub> fixation may, at least partially, have contributed to the higher phytoplankton biomass during the second bloom in our experiment.

Our results indicate an OA-dependent increase in primary production during certain times of the spring-to-summer phytoplankton succession, particularly under NO<sub>3</sub><sup>−</sup> limitation (phase III) being accompanied by a significant increase in picoeukaryotes during this period [21]. With respect to higher trophic levels, OA showed differential growth effects on several pre-dominant mesozooplankton species, though as a whole, the community remained rather unaltered under OA (Alguero et al. *in prep.*). OA led, however, to an increase in the survival rate of herring larvae (being planted in the mesocosms on t63), which could be linked to higher prey abundances (Swaat et al. *in prep.*). Hence, the observed changes in primary production under OA have a high potential to restructure phytoplankton communities in the future coastal North Sea with likely consequences for higher trophic levels.

## Supporting information

**S1 Fig. PAR (μmol m<sup>−2</sup> s<sup>−1</sup>) inside the mesocosms during the time of the experiment.**  
(PNG)

**S1 Table. Raw data of photoacclimation parameters from photosynthesis-irradiance response curves.**  
(XLSX)

## Acknowledgments

We thank the Sven Lovén Centre for Marine Sciences, Kristineberg, for their hospitality and help during the field work. We thank Amy Forsberg-Grivogiannis for her great help during the field work. We also gratefully acknowledge the captain and crew of RV ALKOR for their work transporting, deploying and recovering the mesocosms during cruises AL406 and AL420. This project was funded by the German Federal Ministry of Science and Education (BMBF) in the framework of the BIOACID II project (grant no. FKZ 03F0655A). SW, TE and DBvdW received support from the EU FP7 research infrastructure initiative ‘Association of European Marine Biological Laboratories’ (ASSEMBLE (grant no. 227799), and SW from the Royal Swedish Academy of Sciences.

## Author Contributions

**Conceptualization:** TE BR DBW UJ.

**Formal analysis:** TE BR DBW.

**Investigation:** TE SW.

**Methodology:** TE BR DBW.

**Writing – original draft:** TE.

**Writing – review & editing:** BR DBW LB UJ UR.

## References

1. Wolf-Gladrow DA, Riebesell U, Burkhardt S, Bijma J. Direct effects of CO<sub>2</sub> concentration on growth and isotopic composition of marine plankton. *Tellus B*. 1999; 51: 461–476.
2. Caldeira K, Wickett ME. Anthropogenic carbon and ocean pH. *Nature*. 2003; 425: 365. doi: [10.1038/425365a](https://doi.org/10.1038/425365a) PMID: [14508477](https://pubmed.ncbi.nlm.nih.gov/14508477/)
3. Stocker TF, Qin D, Plattner G-K, Tignor M, Allen SK, Boschung J, et al. (eds.). 2013. In: climate change 2013: the physical science basis. Contribution of working group I to the fifth assessment report of the intergovernmental panel on climate change. Cambridge University Press, USA.
4. Roy T, Bopp L, Gehlen M, Schneider B, Cadule P, Frölicher TL, et al. Regional impacts of climate change and atmospheric CO<sub>2</sub> on future ocean carbon uptake: A multimodel linear feedback analysis. *J. Clim.* 2011; 24: 2300–2318.
5. Sabine CL, Feely RA, Gruber N, Key RM, Lee K, Bullister JL, et al. The oceanic sink for anthropogenic CO<sub>2</sub>. *Science*. 2004; 305: 367–371. doi: [10.1126/science.1097403](https://doi.org/10.1126/science.1097403) PMID: [15256665](https://pubmed.ncbi.nlm.nih.gov/15256665/)
6. Raven JA. Inorganic carbon acquisition by marine autotrophs. *Adv. Bot. Res.* 1997; 27: 85–209.
7. Badger MR, Andrews TJ, Whitney SM, Ludwig M, Yellowlees DC, Leggat W, Price GD. The diversity and coevolution of Rubisco, plastids, pyrenoids, and chloroplast-based CO<sub>2</sub>-concentrating mechanisms in algae. *Can. J. Bot.* 1998; 76: 1052–1071.
8. Young JN, Heureux AMC, Sharwood RE, Rickaby REM, Morel FMM, Whitney SM. Large variation in the Rubisco kinetics of diatoms reveals diversity among their carbon-concentrating mechanisms. *J. Exp. Bot.* 2016;
9. Raven JA. Physiology of inorganic C acquisition and implications for resource use efficiency by marine phytoplankton: relation to increased CO<sub>2</sub> and temperature. *Plant Cell Environ.* 1991; 14: 779–794.
10. Giordano M, Beardall J, Raven JA. CO<sub>2</sub> concentrating mechanisms in algae: mechanisms, environmental modulation, and evolution. *Annu. Rev. Plant. Biol.* 2005; 56: 99–131. doi: [10.1146/annurev.arplant.56.032604.144052](https://doi.org/10.1146/annurev.arplant.56.032604.144052) PMID: [15862091](https://pubmed.ncbi.nlm.nih.gov/15862091/)
11. Rost B, Zondervan I, Wolf-Gladrow D. Sensitivity of phytoplankton to future changes in ocean carbonate chemistry: current knowledge, contradictions and research directions. *Mar. Ecol. Prog. Ser.* 2008; 373: 227–237.
12. Reinfelder JR. Carbon concentrating mechanisms in eukaryotic marine phytoplankton. *Annu. Rev. Mar. Sci.* 2011; 3: 291–315.
13. Raven JA, Beardall J, Giordano M. Energy costs of carbon dioxide concentrating mechanisms in aquatic organisms. *Photosynth Res.* 2014; 121(2–3):111–24. doi: [10.1007/s11120-013-9962-7](https://doi.org/10.1007/s11120-013-9962-7) PMID: [24390639](https://pubmed.ncbi.nlm.nih.gov/24390639/)
14. Burkhardt S, Zondervan I, Riebesell U. Effect of CO<sub>2</sub> concentration on C:N:P ratio in marine phytoplankton: a species comparison. *Limnol. Oceanogr.* 1999; 44: 683–690.
15. Kranz SA, Sültemeyer D, Richter K-U, Rost B. Carbon acquisition by *Trichodesmium*: the effect of pCO<sub>2</sub> and diurnal changes. *Limnol. Oceanogr.* 2009; 54: 548–559.
16. Eberlein T, Van de Waal DB, Brandenburg KM, John U, Voss M, Achterberg EP, et al. Interactive effects of ocean acidification and nitrogen limitation on two bloom-forming dinoflagellate species. *Mar. Ecol. Prog. Ser.* 2016; 543: 127–140.
17. Hutchins DA, Fu F-X, Zhang Y, Warner ME, Feng Y, Portune K, et al. CO<sub>2</sub> control of *Trichodesmium* N<sub>2</sub> fixation, photosynthesis, growth rates, and elemental ratios: Implications for past, present, and future ocean biogeochemistry. *Limnol. Oceanogr.* 2007; 52(4): 1293–1304.



18. Brading P, Warner ME, Smith DJ, Suggett DJ. Contrasting modes of inorganic carbon acquisition amongst *Symbiodinium* (Dinophyceae) phylotypes. *New Phytol.* 2013; 200: 432–442. doi: [10.1111/nph.12379](https://doi.org/10.1111/nph.12379) PMID: [23815769](https://pubmed.ncbi.nlm.nih.gov/23815769/)
19. Schaum E, Rost B, Millar AJ, Collins S. Variation in plastic responses of a globally distributed picoplankton species to ocean acidification. *Nat. Clim. Change.* 2013; 3: 298–302.
20. Riebesell U, Tortell PD. Effects of ocean acidification on pelagic organisms and ecosystems. In: Gattuso J.P., Hansson L. (Eds.), *Ocean Acidification*. Oxford University Press, Oxford, UK, 2011; 99–121.
21. Bach LT, Taucher J, Boxhammer T, Ludwig A, The Kristineberg KOSMOS Consortium, Achterberg EP, et al. Influence of Ocean Acidification on a Natural Winter-to-Summer Plankton Succession: First Insights from a Long-Term Mesocosm Study Draw Attention to Periods of Low Nutrient Concentrations. *PLoS ONE.* 2016; 11(8). e0159068. doi: [10.1371/journal.pone.0159068](https://doi.org/10.1371/journal.pone.0159068) PMID: [27525979](https://pubmed.ncbi.nlm.nih.gov/27525979/)
22. Tiselius P, Kuylensstierna M. Growth and decline of a diatom spring bloom: phytoplankton species composition, formation of marine snow and the role of heterotrophic dinoflagellates. *J. Plankton. Res.* 1996; 18: 133–155.
23. Waite AM, Lindahl O. Bloom and decline of the toxic flagellate *Chattonella marina* in a Swedish fjord. *Mar. Ecol. Prog. Ser.* 2006; 326: 77–83.
24. Steemann Nielsen E. The use of radioactive carbon ( $^{14}\text{C}$ ) for measuring primary production in the sea. *J. Cons. Perm. Int. Explor. Mer.* 1952; 18: 117–140.
25. Riebesell U, Czerny J, Bröckel KV, Boxhammer T, Büdenbender J, Deckelnick M, et al. Technical Note: A mobile sea-going mesocosm system—new opportunities for ocean change research. *Biogeosciences.* 2013; 10(3): 1835–1847.
26. Boxhammer T, Bach LT, Czerny J, Riebesell U. Technical note: Sampling and processing of mesocosm sediment trap material for quantitative biogeochemical analysis. *Biogeosciences.* 2016; 13: 2849–2858.
27. Bender M, Grande K, Johnson K, Marra J, Williams PJ LeB, Sieburth J, et al. A comparison of four methods for determining planktonic community production. *Limnol. Oceanogr.* 1987; 32(5): 1085–1098.
28. Lindahl O. Long-term studies of primary production in the Gullmar fjord, Sweden. In: Skjoldal H. R., Hopkins C., Erikstad K. E., P Leinaas H. [eds.] *Ecology of fjords and coastal waters*. Elsevier Science Publishers, New York. 1995; 105–112.
29. Lindahl O, Belgrano A, Davidsson L, Hernroth B. Primary production, climatic oscillations, and physico-chemical processes: the Gullmar Fjord time-series data set (1985–1996). *ICES J. Mar. Sci.* 1998; 55 (4): 723–729.
30. Egge JK, Thingstad TF, Larsen A, Engel A, Wohlers J, Bellerby RGJ, et al. Primary production during nutrient-induced blooms at elevated  $\text{CO}_2$  concentrations. *Biogeosciences.* 2009; 6: 877–885.
31. Paul AJ, Bach LT, Schulz KG, Boxhammer T, Czerny J, Achterberg EP, et al. Effect of elevated  $\text{CO}_2$  on organic matter pools and fluxes in a summer Baltic Sea plankton community. *Biogeosciences.* 2015; 12: 6181–6203.
32. Sala MM, Aparicio FL, Balagué V, Boras JA, Borrell E, Cardelús C. Contrasting effects of ocean acidification on the microbial food web under different trophic conditions. *Ices. J. Mar. Sci.* 2015;
33. Hopkinson BM, Xu Y, Shi D, McGinn PJ, Morel FMM. The effect of  $\text{CO}_2$  on the photosynthetic physiology of phytoplankton in the Gulf of Alaska. *Limnol. Oceanogr.* 2010; 55(5): 2011–2024.
34. Elser JJ, Bracken MES, Cleland EE, Gruner DS, Harpole WS, Hillebrand H, et al. Global analysis of nitrogen and phosphorus limitation of primary producers in freshwater, marine and terrestrial ecosystems. *Ecol. Lett.* 2007; 10: 1135–1142. doi: [10.1111/j.1461-0248.2007.01113.x](https://doi.org/10.1111/j.1461-0248.2007.01113.x) PMID: [17922835](https://pubmed.ncbi.nlm.nih.gov/17922835/)
35. Moore CM, Mills MM, Arrigo KR, Berman-Frank I, Bopp L, Boyd PW, et al. Processes and patterns of oceanic nutrient limitation. *Nat. Geosci.* 2013; 6: 701–710.
36. Riebesell U, Schulz KG, Bellerby RGJ, Botros M, Fritsche P, Meyerhofer M, et al. Enhanced biological carbon consumption in a high  $\text{CO}_2$  ocean. *Nature.* 2007; 450: 545–548. doi: [10.1038/nature06267](https://doi.org/10.1038/nature06267) PMID: [17994008](https://pubmed.ncbi.nlm.nih.gov/17994008/)
37. Engel A, Borchard C, Piontek J, Schulz KG, Riebesell U, Bellerby R.  $\text{CO}_2$  increases  $^{14}\text{C}$  primary production in an Arctic plankton community. *Biogeosciences.* 2013; 10: 1291–1308.
38. Trimborn S, Lundholm N, Thoms S, Richter K-U, Krock B, Hansen PJ et al. Inorganic carbon acquisition in potentially toxic and non-toxic diatoms: the effect of pH-induced changes in seawater carbonate chemistry. *Physiol. Plant.* 2008; 133: 92–105. doi: [10.1111/j.1399-3054.2007.01038.x](https://doi.org/10.1111/j.1399-3054.2007.01038.x) PMID: [18405335](https://pubmed.ncbi.nlm.nih.gov/18405335/)
39. Wu Y, Gao K, Riebesell U.  $\text{CO}_2$ -induced seawater acidification affects physiological performance of the marine diatom *Phaeodactylum tricornutum*. *Biogeosciences.* 2010; 7: 2915–2923.
40. Hoppe CJM, Holtz L-M, Trimborn S, Rost B. Ocean Acidification decreases the light use efficiency in an Antarctic diatom under dynamic but not constant light, *New Phytol.* 2015; 207: 159–171. doi: [10.1111/nph.13334](https://doi.org/10.1111/nph.13334) PMID: [25708812](https://pubmed.ncbi.nlm.nih.gov/25708812/)